Table 2.—Comparison of differences in fecundity of lobsters as calculated from regression equations derived from Herrick's data and from the Rhode Island analysis

Carapace length (mm)	Total length (inches)	Fecundity			
		Herrick's regression (1)	Rhode Island regression (2)	Difference (1)-(2)	Percent of (2)
70	8.00	5474	5210	+264	5,06
76	8. <u>75</u>	7213	6112	+1101	18.01
85	9.75	10500	8424	+2076	24.64
92	10.50	13695	10569	± 3126	29.57
85 92 98	11.25	16920	12597	14 323	34.31
103	11.75	19958	14602	-∔5356	36.27
107	12.25	22725	16263	+6462	39.73
110	12.50	24934	17631	± 7303	41.42
114	13.00	28110	19530	+8580	43.93
120	13.75	33390	-22628	+10762	47.56
127	14.50	40418	26615	+13803	51.86
133	15.25	47110	30360	+16750	55.17
137	15.75	52030	33060	+18990	57.44
144	16.50	61548	38150	+23398	61.33
149	17.00	68990	42068	+26922	
153	17.50	75408	45378		63.99
158	18.00	84030	49761	+30030	66.17
100	10.00	0.4030	49/01	+34269	68.86

that the variance of the new data is considerably greater than that obtained from Herrick. This is probably due to the fact that only average values of fecundity are available from Herrick's data, and individual values were used for the new study. A test of the hypothesis $\beta_1 = \beta_2$ clearly demonstrated significant differences in the two sample regression coefficients. As stated previously, the difference is apparently due to a bias introduced by Herrick's method of estimating fecundity.

The derived regression equations for the carapace length-fecundity relationships of the American lobster are as follows:

Herrick's data:

$$log_{10}Y(Fecundity) = -2.4505 + 3.3542log_{10}$$

× (carapace length mm)

New data:

$$\begin{array}{c} log_{10}Y(Fecundity) = 1.6017 + 2.8647log_{10} \\ \times \text{ (carapace length mm)} \end{array}$$

The above equations were utilized to calculate fecundity for a range of sizes and to make comparisons of the differences obtained (Table 2). It is clear from an examination of this table that there is a consistent positive difference in the estimated average fecundity for given lengths and that the magnitude of this difference increases with increasing size.

It is suggested from this study that the equation derived from the new estimates of lobster fecundity is more realistic than the one previously utilized based on Herrick's data. The latter estimates of average fecundity are

apparently biased in a positive direction due to the estimation method since the samples were compared at approximately the same period of egg development, and it is assumed that fecundity has not changed since the original samples were taken.

LITERATURE CITED

HERRICK, F. H. 1911. Natural History of the American Lobster. Bull. U. S. Bur. Fish. 29: 149-408. HUGHES, J. T. 1965. Comm. of Massachusetts, Div.

Marine Fish., Annual Rept.: 18-28. Saila, S. B., and J. M. Flowers. 1966. tion study of sex ratios and regulation effects with the American Lobster, Homarus americanus. Proc. Gulf and Caribbean Fisheries Institute, 18th Ann. Session, Nov., 1965: 66-78. SNEDECOR, G. W., AND W. G. COCHRAN. 1967. Sta-

tistical methods. 6th ed. xvi + 593 pp.

S. B. SAILA J. M. FLOWERS

Graduate School of Oceanography University of Rhode Island Kingston, Rhode Island 02881

J. T. Hughes

Massachusetts Division of Marine Fisheries State Lobster Hatchery and Research Station Vineyard Haven, Massachusetts 02568

The Effects of a Rotenone Treatment on the Insect fauna of a California Stream

In the late summer of 1963, the California Department of Fish and Game embarked upon a "rough fish" control program of the major tributaries of the Russian River, north

Although chemical treatment of streams to remove unwanted fish species is a common fisheries management practice (see Burns 1966, and Meyer 1966 for reviews), this practice has been subject to justifiable criticism (Hubbs, 1963). One question that has been raised concerns the possible adverse effects of these treatments on the non-target invertebrate fauna of the streams so affected. It is this point that forms the basis for this report.

One of the streams selected for treatment under this project was Robinson Creek, Mendocino County. Robinson Creek is about 10 miles in length and drains a fairly extensive mountainous watershed from the eastern slope of the central Coast Range. As is typical of most streams in this region, water volume varies from several thousand cubic feet per second during winter storms to mere trickles in the summer; sections of many of the streams go dry or underground during the dry season.

In the general area of concern here, the creek flows through a canyon for about 3 miles. The upper part of the canyon is steep sided and narrow, gradually opening out as it approaches the valley floor of the Russian River. This is accompanied by a decrease in the stream gradient. The substrate varies locally with the stream gradient from bedrock to silty gravel in the quiet pools. Water-worn rocks and boulders characterize much of the stream bed. The canyon walls are well vegetated with many of the trees and shrubs characteristic of the broad sclerophyll association in this region.

The treatment plan for Robinson Creek was to apply rotenone to the stream below a point 4½ miles above its mouth. It was at approximately this point where visual observation indicated an upstream predominance of trout

and a downstream predominance of suckers, western roach, sculpins, squawfish, and larval lampreys. It was also at approximately this point where the canyon began to open up gradually and the stream gradient decreased.

The data presented here represent an attempt to determine the overall effect of the rotenone treatment of Robinson Creek on the predominant insect groups. Neither time nor facilities permitted identification of these organisms beyond the ordinal of familial level.

METHODS AND PROCEDURES

The procedural plan that appeared best adapted to the objectives of this study was to follow the population levels of the major insect groups subsequent to treatment in the treated and untreated zones. Consequently, sampling stations were established within a few hundred yards above and below the point where the treatment began:

Two major habitat types were sampled: the pools and the riffles. Although the pools appeared fairly similar throughout, the riffles varied in gradient from what might be typically regarded as a riffle of fairly low gradient to small cascades where the water flowed over, around, and between boulders in the stream bed. Between 2 and 4 samples were taken at irregular intervals from each of the habitat types from both the treated and untreated zones as water conditions permitted from 3 days after the treatment until the following spring.

The sampling method originally considered for use in this study was based upon the Surber sampler. Due, however, to the rocky character of the stream bed, and the limited amount of water in the creek during the late. summer, this method proved generally unsatisfactory for purposes of this study. Instead, a method was employed which took advantage of the rocks that formed so much of the substrate. At each station 5 rocks were carefully removed from the stream bed and washed into a fine-mesh (32 squares/inch) sieve. The organisms and other residue so obtained were then washed into pint jars. Every attempt was made to sample rocks of roughly the same size from all stations. Although this was not possible in all cases, most

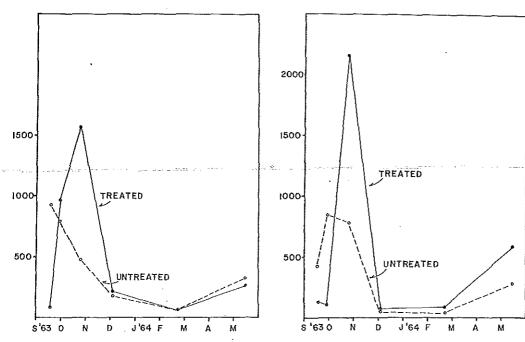


FIGURE 1.—Total insect fauna: riffle samples.

FIGURE 2.—Total insect fauna: pool samples.

of the rocks sampled were in the 2 to 3 pound range. When it became necessary to sample larger or smaller sizes, compensations were made to adjust for significant differences in surface area.

Robinson Creek was treated on September 12, 1963, by the California Department of Fish and Game with the authors as observers. The project plan called for dispensation into the water of *Pronoxfish* "at a concentration level of not less than 0.050 active ingredient rotenone."

RESULTS

The predominant groups of insects encountered in Robinson Creek during the course of this study were representatives of the orders Trichoptera, Ephemeroptera, and Diptera. Although the caddisflies and mayflies were not broken down in this report beyond the ordinal level, they were primarily represented in the samples by leptocerids and rhyacophilids, and heptageniids respectively. The Diptera were broken down to the family with simuliids and chironomids (tendipedids) predominating in this group.

As would be expected, the effects of the treatment became apparent within a few min-

utes after the material was applied to the water. A small mesh net placed below a riffle yielded large numbers of dead and dying aquatic invertebrates as well as fish. The first sample recorded was taken on September 15, 1963, 3 days following the treatment.

The effects of the poisoning and the subsequent recovery of the total insect fauna in the treated portion of Robinson Creek can be compared with the control, or untreated zone, in Figures 1 and 2. As can be seen in both of these figures, there was a rapid and explosive resurgence of the insect fauna in both the habitat types sampled in the treated zone following initial near annihilation. With the onset of winter, however, population levels in the treated zone of the stream had declined to a point commensurate with population levels in the untreated zone. This trend continued with the riffle fauna until the next spring (Figure 1). The pool fauna (Figure 2), however, appeared to carry populations in the treated zone approximately double to that in the untreated zone.

The major components of the riffle fauna were found to be larvae of blackflies (Figure

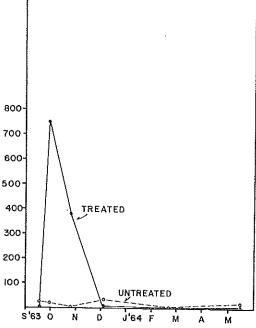


FIGURE 3.—Diptera: Simuliidae: riffle samples.

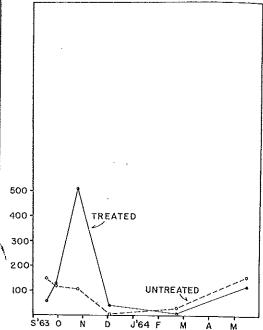


FIGURE 4.-Trichoptera: riffle samples.

3), caddisflies (Figure 4), and mayflies (Figure 5). Chironomid midges by far predominated in the pools (Figure 6), with trichopters and ephemerids contributing approximately 10% to the total insect fauna in this habitat.

The first major group to make a comeback in the treated riffles was the simuliids (Figure 3). Within two weeks subsequent to treatment, these organisms had virtually taken over all the available attachment space in the moving water. Although in streams blackfly larvae are most generally associated with fast moving water, they were everywhere at this time. Significantly, they were all early instars, indicating recent reproduction rather than drift from the untreated zone above. Within six weeks after the treatment, the simuliid population had dropped significantly. The caddisfly and mayfly larval populations, however, had erupted by this time (Figures 4 and 5). These were also represented by early instars, although a few large individual caddisfly larvae indicated an incomplete kill or drift from the upstream untreated zone.

In the pools, the chironomid midge larvae also erputed in the treated zone approximately

6 weeks after the treatment (Figure 6). These were also almost exclusively early instars.

There were several other major insect groups encountered during this study, although collectively they represented less than 5% of the total insect fauna sampled. Of these, the Plecoptera were found to be most abundant. These were also virtually annihilated in the treated zone, but had attained densities commensurate with those of the untreated zone by the following spring. Of the remaining groups, too few individuals were collected to evaluate. These included species of Odonata, Coleoptera (Psephenidae), Lepidoptera (Pyralidae: Nymphulinae), and Diptera (Tipulidae, Stratiomyidae, Ceratopogonidae, Tabanidae, and Blephariceridae). Other organisms included aquatic mites, copepods, ostracods, nematodes, Planaria, leeches, annelid worms, and snails.

CONCLUSIONS

From an entomological standpoint, the most interesting aspect of this study concerned the great resurgence of the insect fauna shortly after their initial near annihilation in the treated zone. This was particularly interesting

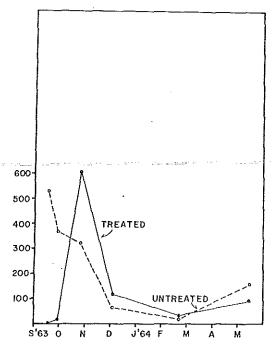


FIGURE 5.- Ephemeroptera: riffle samples.

in the case of the simuliids. As alluded to above, in environments such as Robinson Creek, these organisms are associated with the swiftest moving water. In view of the observations made here, it would appear that blackfly larvae may be limited in their ecological distribution by predation pressure rather than by any highly specific environmental needs, such as particularly fast moving water. As mentioned previously, when the Simuliidae returned after the treatment, they were attached to rocks, twigs, leaves, aquatic vegetation, and any other stationary objects in the stream bed. A few were even found in the pools. The elimination of the fish and other potential predators from this portion of the stream may account in part for the unusual abundance and survival of these organisms during the several weeks subsequent to the treatment.

On the basis of the information obtained in this limited study, it would appear that the application of rotenone to a stream for the purpose of reducing unwanted fish species has little lasting effect upon the non-target insect fauna of significance as fish forage. Caution,

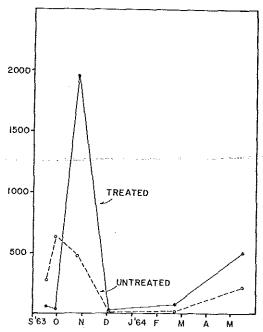


FIGURE 6.—Diptera: Chironomidae: pool samples.

however, should be used in projecting these results to other instances of stream poisoning. In the case of Robinson Creek, only a portion of the stream was treated; this permitted repopulation from the untreated zone. Also, the timing of such a treatment could be very important in the case of those species that produce but one generation per year, or those with highly synchronous emergence times. If the treatment occurred during that time when the entire population was in the larval stage, repopulation could be significantly delayed.

Other points to be considered in the evaluation of these results concern possible biases due to the sampling methods, and possible species shifts that went undetected due to the broad identification procedures. It was unfortunate that a more detailed study was not possible in this case. It is entirely possible that had the identifications been carried to the specific level, that species shifts in predominance might have been detected.

ACKNOWLEDGMENT

We wish to express our sincere thanks to Mrs. S. G. Herman for preparation of the figures.

LITERATURE CITED

Burns, J. W. 1966. Rough fish management. In:
Inland Fisheries Management (A. Calhoun,
Editor). California Dept. Fish and Game publication: 492-498.
Hubbs, Clark. 1963. An evaluation of the use of a
rotenone as a means of "improving" sport fishing
in the Concho River, Texas. Copeia 1963: 199203.

Meyer, F. A. 1966. Chemical control of undesirable fishes. In: Inland Fisheries Management

(A. Calhoun, Editor). California Dept. Fish and Game publication: 498-510.

S. F. Cook, JR.

Department of Zoology University of California Davis, California

R. L. MOORE

Department of Fish and Game Ferry Building San Francisco, California 94111